



Effects of Highly Active Antiretroviral Treatment on Liver and Kidney Functions

Simon Bannison Bani^{1*}, Christian Obirikorang², Kwabena O. Danquah³,
William K. B. A. Owiredu², Lawrence Quaye¹, Samuel A. Sakyi², Yussif Adams¹,
Peter Paul M. Dapare¹, Moses Banyeh¹ and Barnabas B. N. Gandau⁴

¹Department of Biomedical Laboratory Science, School of Allied Health Sciences, University for Development Studies, Tamale, Ghana.

²Department of Molecular Medicine, School of Medicine and Dentistry, College of Health Sciences, Kwame Nkrumah University of Science and Technology, Kumasi, Ghana.

³Department of Medical Laboratory Technology, Faculty of Allied Health Sciences, College of Health Sciences, Kwame Nkrumah University of Science and Technology, Kumasi, Ghana.

⁴School of Medicine and Health Sciences, University for Development Studies, Tamale, Ghana.

Authors' contributions

This work was carried out in collaboration among all authors. Authors WKBAO, CO, SAS and KOD conceived and designed the experiments. Authors SBB, YA, CO and KOD performed the experiments. Authors LQ, SBB, YA, CO and KOD analyzed the data. Authors SBB, YA, CO, SAS and KOD contributed reagents/materials/analysis tools and other logistics. The manuscript was written and edited by authors LQ, SBB, EA, WKBAO and KOD. Interpretation of results and appraisal of manuscript by authors PPMD, EA and BBNG. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/ISRR/2020/v9i230109

Editor(s):

(1) Dr. Kailash Gupta, National Institute of Allergy and Infectious Diseases, USA.

Reviewers:

(1) Jane Mathew, NGSM Institute of Pharmaceutical Sciences (Constituent College of NITTE Deemed to be University), India.

(2) M. B. Ravikumar, Mandya Institute of Medical Sciences, India.

Complete Peer review History: <http://www.sdiarticle4.com/review-history/59518>

Original Research Article

Received 27 May 2020
Accepted 03 August 2020
Published 15 August 2020

ABSTRACT

Aim: This study assesses the effects of HAART on liver and renal functions in HIV infected individuals on HAART.

Study Design: Cross sectional study.

Place and Methods: This study was conducted in Tamale, Ghana from August, 2015 to November 2017.

*Corresponding author: Email: bsbannison@yahoo.co.uk, bsbannison@uds.edu.gh;

Methodology: A total of 300 HIV infected participants with ages ranging from 19 to 79 years who have been administered with HAART for at least 6 months were recruited. Pre-HAART administration (baseline) demographic and clinical information, with initial liver and renal function test results were retrieved from the medical records of the participants present at the ART center. Post HAART administration blood sample (5 mLs) was taken from each participant into a gel separated vacutainer tube, allowed to clot and spun at 3000 rpm for 3 minutes to produce serum. The product (serum) was used for liver and renal function test analysis using a fully automated chemistry analyser (Vital Scientific Selectra Flexor XL).

Results: Of the study population, 72% were administered with AZT/3TC/EFV, 13% with AZT/3TC/NVP, 6.7% with TDF/3TC/LPV/r and TDF/3TC/NVP, 1% with AZT/3TC/EFV while 0.7% were administered with TDF/FTC/EFV. The following parameters were significantly increased post HAART administration; ALT (25.53 ± 16.90 to 30.87 ± 19.28 U/L), ALP (163.7 ± 141.0 to 215.2 ± 143.4 U/L), GGT (37.27 ± 25.21 to 53.19 ± 41.71 U/L), Total protein (73.97 ± 17.08 to 82.31 ± 11.62 g/L), Albumin (38.02 ± 9.331 to 41.01 ± 7.471 g/L), Globulin 38.02 ± 15.71 to 42.79 ± 25.20 (g/L). There were however significant reductions in Total bilirubin (12.13 ± 10.85 to 9.434 ± 4.560 $\mu\text{mol/L}$), Direct bilirubin (6.616 ± 5.770 to 4.184 ± 2.806 $\mu\text{mol/L}$), (Creatinine 73.19 ± 36.13 to 63.14 ± 27.14 $\mu\text{mol/L}$) and Urea (3.515 ± 2.552 to 3.011 ± 1.274 mmol/L).

Conclusion: HAART improves renal function, induces elevation in liver enzymes, stimulates the production of plasma proteins and reduces serum bilirubin concentration.

Keywords: Highly active antiretroviral therapy; HIV infection; LFT; RFT; HIVAN.

ABBREVIATIONS

Short term HAART users = Participants who have been on HAART for less than or equal to 52 months; Medium term HAART users = Participants who have been on HAART for more than 52 months but less than or equal to 104 months; Long term HAART users = Participants who have been on HAART for more than 104 months.

1. INTRODUCTION

HIV infection leading to immune deficiencies which allow fatal opportunistic infections has been a major public health concern of the 21st century. Globally, 36.9 million people are living with HIV whereas 21.7 million are on antiretroviral therapy (UNAIDS, 2017). About 35.4 million people have died of AIDS related illnesses since the start of the epidemic (UNAIDS, 2017). Research has shown significant renal impairment among HAART naïve HIV infected individuals [1]. HIV associated nephropathy (HIVAN) is the most common kidney disease amongst people living with HIV [2,3]. An estimated 5.9% of HAART naïve people living with HIV present with acute renal failure [4]. Renal disorders ranging from electrolyte and fluid imbalances to end stage renal disease (ESRD) is evident in all stages of HIV infection [5]. In caring for individuals infected with HIV on HAART, managing liver and renal diseases remain very important. HAART induced liver disease is the

most common non HIV associated cause of death amongst people living with HIV on HAART, causing about 14 to 18% of deaths in this population and about 50% of all hospitalized HIV infected individuals [6]. HAART related liver toxicity is one of the most important adverse events reported amongst people living with HIV [7]. It usually presents as mild abnormalities in liver function parameters to very pronounced liver failure [8].

HAART is a multi-drug combination regimen targeted to reduce HIV viral replication to undetectable concentrations and keep HIV viral replication suppressed for years in infected individuals. HAART has been able to significantly reduce mortality and morbidity associated with AIDS [9]. The advent of HAART has significantly improved the life expectancy of HIV infected individuals who are on it [3]. Over time however, the use of HAART results in hepatotoxicity and nephrotoxicity [10]. Renal dysfunction has been associated with tenofovir disoproxilfumarate which accumulates in the proximal tubule of the kidney [11]. HAART induces functional stress especially mitochondrial injury. Nevirapine and efavirenz may lead to hypersensitivity syndrome reactions that may lead to hepatotoxicity which leads to liver necrosis and death [12]. Reisler [13] reported the rate of hepatotoxicity associated with nucleotide reverse transcriptase inhibitors (NRTIs) as 12%, which brings to light the difficulty in managing HAART associated liver

and renal pathologies [14]. The aim of this study was to assess the effects of HAART on liver and renal functions in HIV infected individuals on HAART.

2. MATERIALS AND METHODS

2.1 Study Design

This was a cross sectional study carried out from August 2015 to November, 2017.

2.2 Study Population

A total of 300 HIV infected participants with ages ranging from 19 to 79 years have been administered with HAART for at least 6 months were recruited at the Tamale Teaching Hospital, Savelugu District Hospital, Tamale West and Central hospitals.

2.3 Data Collection

Pre-HAART administration (baseline) demographic and clinical information, with initial liver and renal function test results were retrieved from the medical records of the participants present at the ART centers. Post HAART information such as age, weight, HAART type and duration of HAART use were recorded using questionnaire which had been pre-tested among 10 HIV infected individuals administered with HAART to clear possible ambiguity and difficulty in answering the questions. Data from the pre-tested questionnaires were however not included in the results analysis.

Post HAART blood sample (5mLs) was taken from each participant into a gel separated vacutainer tube, allowed to clot, spun at 3000rpm for 3 minutes to produce serum which was used to perform liver and renal function tests using a fully automated chemistry analyzer, Vital Scientific Selectra Flexor XL. The HAART comprised two Nucleotide Reverse Transcriptase Inhibitor (NRTI) plus one Non-nucleotide Reverse Transcriptase Inhibitor (NNRTI) or two NRTIs and a Protease Inhibitor (PI). The participants were further stratified into short term (<52 months), medium term (≥ 52 but ≤ 104 months), long term (>104 months) based on the duration of HAART.

2.4 Statistical Analysis

Data was entered into Microsoft excel 2016 and exported to GraphPad prism version 6.0 (www.graphpad.com) for analysis. Data was presented as number, percentages, means and standard deviation. Means were compared between groups using student paired t-test and ANOVA. A $p < 0.05$ was considered statistically significant.

3. RESULTS AND DISCUSSION

3.1 Demographic Characteristics of the Study Population

According to the demographic data of the study population as captured in Table 1, A total of 300 subjects were recruited and were grouped into cases and controls. In all, 217 (72.3%) were classified as cases, while 83 (27.7%) were classified as controls. The cases were further classified into short-term, medium-term and long-term HAART users depending on the duration of HAART usage. Participants administered with HAART for 52 months or less months were classified as short-term HAART users, those who have been administered with HAART for more than 52 months but less than or equal to 104 months were classified as medium-term HAART users, while more 104 months were classified as long-term HAART users.

3.2 Distribution of HAART in the Study Population

The antiretrovirals administered to the study participants in this study included Zidovudine (AZT), Lamivudine (3TC), efavirenz (EFV), Nevirapine (NVP), tenofovir (TDF), lopinavir (LPV), NVP, emtricitabine (FTC) and ritonavir (r). HAART was administered as triple combinations as follows; AZT/3TC/EFV, AZT/3TC/NVP, TDF/3TC/EFV, TDF/3TC/LPV, TDF/3TC/LPV/r, TDF/3TC/NVP, TDF/FTC/EFV. Of all the participants, 3/300 (1%) were administered with AZT/3TC/EFV, 39/300 (13.0%) were administered with AZT/3TC/NVP, 216/300 (72%) of the participants were administered with TDF/3TC/EFV, 20/300 (6.7%), 20/300 (6.7%) and 2/300 (0.7%) were administered with TDF/3TC/LPV/r, TDF/3TC/NVP and TDF/FTC/EFV respectively.

3.3 Comparison of Liver and Renal Function Parameters Pre and Post HAART

There was an insignificant increase in the mean plasma concentration of AST, from 32.19 ± 21.68 U/L pre-HAART administration to 34.88 ± 26.70 U/L post HAART ($p = 0.6110$). The mean plasma ALT concentration increased significantly from 25.53 ± 16.90 U/L pre-HAART

administration to 30.87 ± 19.28 U/L post HAART ($p = 0.0440$). Also, the mean plasma ALP increased significantly from 163.7 ± 141.0 U/L to 215.2 ± 143.4 U/L post HAART ($p = 0.0002$). Mean plasma GGT concentration increased significantly from 37.27 ± 25.21 U/L to 53.19 ± 41.71 U/L at post HAART ($p < 0.0001$). Table 2 summarises the routine liver and renal function parameters pre and post HAART.

Table 1. Demographic characteristics of the study population

Variables	Pre-HAART (n=300)	Post-HAART (n=300)
Age (years)*	35.4 ± 9.4	39.7 ± 10.0
Weight (Kg)*	58.8 ± 12.8	64.3 ± 25.6
Age Group (years)		
10—19	10 (3.3%)	3 (1%)
20-29	76 (25.3%)	36 (12%)
30-39	125 (41.7%)	120 (40%)
40-49	64 (21.3%)	96 (32%)
50-59	20 (6.7)	34 (11.3%)
60-69	5 (1.7%)	13 (4.3%)
70-79	0	1 (0.3%)
Gender		
Male	58 (19.3%)	58 (19.3%)
Female	242 (80.7%)	242 (80.7%)
HAART Duration (Months)		
Short-term	0	176 (58.7%)
Medium-term	0	96 (32%)
Long-term	0	28 (9.3%)

Data are presented as frequencies and percentages, * - comparison between pre and post HAART, with p-value of paired t-test < 0.001 . P-values 0.05 were considered significant

Table 2. Liver function tests parameters pre and post HAART

Parameters	Pre-HAART	Post-HAART	p - value
AST (U/L)	32.19 ± 21.68	34.88 ± 26.70	0.6110
ALT (U/L)	25.53 ± 16.90	30.87 ± 19.28	0.0440
ALP (U/L)	163.7 ± 141.0	215.2 ± 143.4	0.0002
GGT (U/L)	37.27 ± 25.21	53.19 ± 41.71	0.0001
Total protein (g/L)	73.97 ± 17.08	82.31 ± 11.62	< 0.0001
Albumin (g/L)	38.02 ± 9.331	41.01 ± 7.471	< 0.0001
Globulin (g/L)	38.02 ± 15.71	42.79 ± 25.20	0.0426
Total bilirubin ($\mu\text{mol/L}$)	12.13 ± 10.85	9.434 ± 4.560	0.0043
Direct bilirubin ($\mu\text{mol/L}$)	6.616 ± 7.570	4.184 ± 2.806	0.0002
Indirect bilirubin ($\mu\text{mol/L}$)	12.13 ± 10.85	9.434 ± 4.560	0.0043
Creatinine	73.19 ± 36.13	63.14 ± 27.14	0.0050
Urea	3.515 ± 2.552	3.011 ± 1.274	0.0097

Data are presented as mean \pm SD, shows the comparison of liver function test parameters between HAART users at pre-HAART and Post HAART, p-value < 0.05 was considered statistically significant. AST: Aspartate Aminotransferase, ALT: Alanine Aminotransferase, ALP: Alkaline Phosphatase, GGT: Gamma Glutamyl Transferase

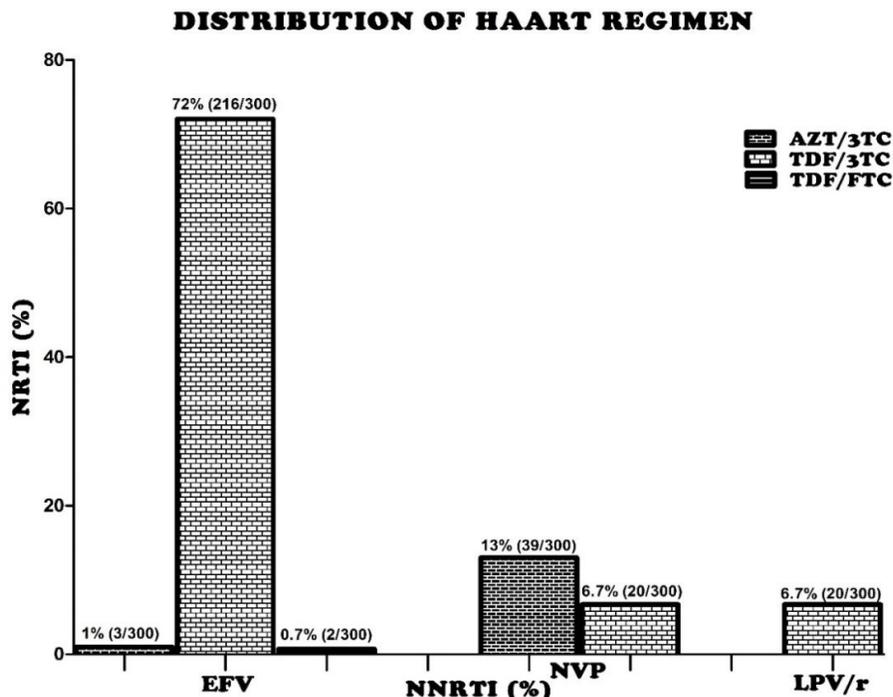


Fig. 1. Shows the percentage distribution of the various types of HAART regimen administered at the ART centers

Table 3. One-way ANOVA of liver and renal function parameters based on the duration of HAART usage

Parameter	Duration of HAART			F test	p value
	Short term	Medium term	Long term		
AST	36.71 ± 27.98	30.07 ± 13.03	30.33 ± 12.09	2.853	0.0593
ALT	32.57 ± 21.29	27.15 ± 12.82	32.68 ± 22.80	2.525	0.0819
GGT	46.82 ± 29.17	45.17 ± 20.87	40.49 ± 15.13	0.6195	0.4923
ALP	219.5 ± 143.6	201.3 ± 153.3	299.0 ± 106.1	2.023	0.5389
Total Protein	82.72 ± 12.22	81.45 ± 10.27	81.87 ± 13.68	0.3658	0.6940
Albumin	40.23 ± 6.930*	42.72 ± 8.257	40.79 ± 7.273	3.433	0.0336
Globulin	44.82 ± 31.74	38.32 ± 9.405	42.93 ± 10.88	1.981	0.1398
Total bilirubin	9.234 ± 4.729	9.630 ± 4.472	11.7 ± 8.4	2.600	0.0760
Direct bilirubin	4.024 ± 2.786	4.299 ± 2.804	5.265 ± 2.824	2.071	0.1280
Indirect bilirubin	5.181 ± 3.158	4.299 ± 2.804	5.324 ± 2.490	2.859	0.0590
Creatinine	62.36 ± 27.26	64.23 ± 28.31	64.17 ± 22.57	0.1627	0.8500
Urea	3.086 ± 1.754	3.198 ± 1.374	3.336 ± 1.316	0.3501	0.7049

Data are presented as mean ± SD, shows One-way ANOVA of Liver and Renal Function Parameters in short, * represents significant comparison between Short-term and Medium-term HAART users. p-value <0.05 was considered statistically significant

3.4 Variations in Liver and Kidney Function Parameters with Duration of HAART

Table 3 shows how liver and kidney function parameters varied among short, medium- and long-term HAART users. Of all the parameters

considered, only albumin varied significantly from 40.23 ± 6.930g/dL in short-term HAART users to 42.72 ± 8.257g/dL in the medium HAART users. Table 3 summarises the comparisons of liver and renal function parameters between short, medium- and long-term HAART users.

3.5 Discussion

The advent and administration of HAART comes along with HAART induced hepatotoxicity. As the life expectancies of HIV infected individuals improved, non-AIDS diseases became increasingly very important cause of morbidity and mortality in the HIV infected population [15]. Liver related diseases have become rampant in HIV infected populations [16,17]. Liver related diseases account for 13-18% of all mortality and one of the leading causes of non-AIDS related cause of death in HIV infected populations. The mechanisms involved in the cause of liver disease in HIV infected individuals on HAART include senescence nodular regenerative hyperplasia, systemic inflammation, cytotoxicity, immune related injury, mitochondrial injury, Oxidative stress, gut microbial translocation, lipotoxicity etc.

This study recorded elevated alanine aminotransferase (ALT), alkaline phosphatase (ALP) and gamma glutamyl transferase (GGT) post HAART administration. Shiferaw [18] and Owiredu [19] also recorded elevated ALT, ALP and GGT post HAART administration. These liver enzymes were associated with hepatocytes and were released into the plasma when the virus compromised the integrity of the hepatocytes. The elevated liver enzymes may be as a result of the toxic effects of HAART or direct inflammation on hepatocytes which increased the permeability of the cell and mitochondrial membranes [20,21]. The elevations in liver enzymes were reported to return to normal after the discontinuation of HAART [22,23]. NRTI incorporation terminated viral DNA maturation, inhibited mitochondrial DNA polymerase, damaged mitochondrial, depleted and impaired cellular respiration [24]. Studies by Kontorinis and Dieterich [24] showed that severe NRTIs induced mitochondrial toxicity which resulted in hepatomegaly and steatosis. NNRTIs, especially efavirenz and Nevirapine also increased the concentration or activity of other co-administered antiretroviral agent, thereby increased their toxicity effect on hepatocytes [25]. Kontorinis and Dieterich (24) recorded that, the activity of ritonavir was increased 20-fold when co-administered with saquinavir. Individuals on PIs recorded more incidences of hepatotoxicity than those on NRTIs [26]. The use of ritonavir recorded 48% hepatotoxicity [22]. It was noted that super therapeutic concentrations of HAART in serum altered its metabolism by the various isoforms of P450 systems [27]. Sulkowski (22)

reported that increasing ritonavir exposure by 40% resulted in a mild hepatic impairment [28,29]. The statement 'the dosage of indinavir should be reduced to 600 mg every 8 hours in patients with mild-to-moderate hepatic insufficiency due to cirrhosis' was contained in the prescriber information in the USA. This showed that the administration of HAART resulted in hepatotoxicity [30] as observed in this study.

This study recorded significant increases in serum total protein post HAART administration. Serum total protein, albumin and globulins were all elevated post HAART administration. The elevation in albumin concentration was believed to be due to the general improvement in the health status of the individual. Globulins and total proteins were higher post haart administration. This confirms the presence of an infection which is able to stimulate the body's immune response leading to the production of antibodies which are immunoglobulins (plasma proteins) [31,32]. This finding concurs with the report of Ngala [33] who also reported elevations in total protein, albumin and globulins in HIV infected individuals post HAART administration. HAART induced improvement in immune status helps in the prevention of opportunistic infection which impairs humoral immune response e.g. pneumococcal pneumonia [34]. According to Ngala [33] as the general immune status of the study population improved, more immune proteins such as antibodies against the disease and opportunistic infections were produced by the body. Improvements in B-cell number, coupled with increased B cell polyclonal activation led to increased serum globulin production post HAART administration. The serum globulins consisted of alpha globulins-1 (10%), alpha globulins-2 (30%), beta globulins (34%), gamma globulins (46%). Picker [35] reported increases in all fractions of globulins post HAART administration.

In this study, total, direct and indirect bilirubins were all reduced post HAART. This is consistent with findings of Mahato [36] who also reported reductions in bilirubin post HAART administration. However, this finding differs from the observations made by Sulkowski [26] and Ayelagbe [37] who found increases in bilirubin post HAART administration. They attributed the increases in bilirubin post HAART to drug mediated impairment of bilirubin conjugation by the bilirubin UDP-glucuronyltransferase which led to decreased conjugation and excretion of

bilirubin [38]. A decrease in bilirubin concentration post HAART administration as observed in this study may be a manifestation of an improvement in renal function which facilitated the rapid excretion of conjugated bilirubin from the body and an absence of opportunistic infections.

This study reported reductions in serum creatinine and urea post HAART administration. This observation contradicts the findings of Obirikorang [3] but corroborated the findings of Betjes and Verhagen [39] who also recorded improvements in serum creatinine and urea concentrations post HAART administration. The improvements in renal function noted in this study basically resulted from HAART's suppression of viral replication and a general improvement in the health status of the study population. The pathogenesis of HIV associated renal disease (HIVARD) includes the direct cytotoxic influence of HIV on renal cells. According to Schwartz [40], the invasion of renal cells by HIV caused structural changes particularly in podocytes which led to loss of differentiation, atrophy, cytolysis and loss of contact inhibition, collapsed glomerulosclerosis and proteinuria. The tubular cells also dedifferentiated, atrophied and underwent apoptosis, causing cystic dilatation [41]. Extensive fibrosis of the interstitium has been reported, with varying degrees of infiltration by mononuclear cells [41,42]. Renal insufficiency mostly occurs in advanced HIV infections with high viral load [42]. The results of this study show HAART-associated improvement in function. Considering the renal pathogenesis of HIVARD as aforementioned, HAART lowering of viral load may be considered best treatment for HIVARD. Monotherapy with a reverse transcriptase inhibitor like AZT was associated with a better renal prognosis especially early in the course of the disease [39,42]. The findings of this study suggest that commencement of HAART before renal complications develop may be the reason for the improved renal function noticed in this study.

4. CONCLUSION AND RECOMMENDATION

HAART administration improves renal function. HAART also causes elevations in liver enzymes, increases the production of plasma proteins and reduces bilirubin, probably as a result of increased excretion due to improvements in renal function.

CONSENT

Written informed consent was obtained from all participants for publication of this case control study.

ETHICAL APPROVAL

This study was approved by the Committee for Human Publication and Research Ethics (CHRPE) of the Kwame Nkrumah University of Science and Technology, Kumasi Ghana in a letter referenced CHRPE/AP/367/16. The study team also obtained institutional approval in a letter referenced GHS/NR/18-O. Written informed consent was obtained from all participants before recruiting them into the study. Consent form was given to each participant to sign or thumb-print and confidentiality was assured.

ACKNOWLEDGEMENTS

The authors wish to express their profound gratitude to all the staff and HIV infected individuals on HAART at the Sexually Transmitted Infections (STI) clinic of the Tamale Teaching Hospital, Tamale Central Hospital, Tamale West Hospital and the Savelugu District Hospital who voluntarily participated in the research.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Onodugo OD, Chukwuka C, Onyedum C, Ejim E, Mbah A, Nkwo P, et al. Baseline renal function among antiretroviral therapy-naive, hiv-infected patients in Southeast Nigeria. *Journal of the International Association of Providers of AIDS Care (JIAPAC)*. 2014;13(5):476-80.
2. Ragwa FG, Waithaka SK, Ouma JH. Comparative renal function tests between HIV patients on and not on antiretrovirals and HIV negative individuals at Nyeri Provincial General Hospital, Nyeri County, Kenya. *Int J Adv Multidiscip Res*. 2017; 4(3):51-8.
3. Obirikorang C, Osakunor DNM, Ntaadu B, Adarkwa OK. Renal function in Ghanaian HIV-infected patients on highly active antiretroviral therapy: A case-control study. *PLoS one*. 2014;9(6):e99469.

4. Franceschini N, Napravnik S, Eron Jr JJ, Szczech LA, Finn WF. Incidence and etiology of acute renal failure among ambulatory HIV-infected patients. *Kidney international*. 2005;67(4):1526-31.
5. Jha V, Garcia-Garcia G, Iseki K, Li Z, Naicker S, Plattner B, et al. Chronic kidney disease: Global dimension and perspectives. *The Lancet*. 2013;382(9888):260-72.
6. Price JC, Thio CL. liver Disease in the Hiv-infected Individual. *Clinical Gastroenterology and Hepatology*. 2010; 8(12):1002-12.
7. Abraham AG, Althoff KN, Jing Y, Estrella MM, Kitahata MM, Wester CW, et al. End-stage renal disease among HIV-infected adults in North America. *Clinical Infectious Diseases*. 2014;60(6):941-9.
8. Althoff KN, McGinnis KA, Wyatt CM, Freiberg MS, Gilbert C, Oursler KK, et al. Comparison of risk and age at diagnosis of myocardial infarction, end-stage renal disease, and non-AIDS-defining cancer in HIV-infected versus uninfected adults. *Clinical Infectious Diseases*. 2014;60(4):627-38.
9. Zhang F, Dou Z, Yu L, Xu J, Jiao JH, Wang N, et al. The effect of highly active antiretroviral therapy on mortality among HIV-infected former plasma donors in China. *Clinical infectious diseases*. 2008; 47(6):825-33.
10. Peters PJ, Moore DM, Mermin J, Brooks JT, Downing R, Were W, et al. Antiretroviral therapy improves renal function among HIV-infected Ugandans. *Kidney international*. 2008;74(7):925-9.
11. Cihlar T, Ho ES, Lin DC, Mulato AS. Human renal organic anion transporter 1 (hOAT1) and its role in the nephrotoxicity of antiviral nucleotide analogs. *Nucleosides, Nucleotides and Nucleic Acids*. 2001;20(4-7):641-8.
12. Reust CE. Common adverse effects of antiretroviral therapy for HIV disease. *American Family Physician*. 2011;83(12).
13. Reisler R, Liou S, Servoss J, Robbins G, Theodore D, Murphy R, et al, editors. Incidence of hepatotoxicity and mortality in 21 adult antiretroviral treatment trials. Program and abstracts of The 1st IAS Conference on HIV Pathogenesis and Treatment; 2001.
14. Reisler R, Liou S, Servoss J, Robbins G, Theodore D, Murphy R, et al, Editors. Incidence of hepatotoxicity and mortality in 21 adult antiretroviral treatment trials. Program and abstracts of The 1st IAS Conference on HIV Pathogenesis and Treatment; Buenos Aires, Argentina; 2001.
15. Palella Jr FJ, Delaney KM, Moorman AC, Loveless MO, Fuhrer J, Satten GA, et al. Declining morbidity and mortality among patients with advanced human immunodeficiency virus infection. *New England Journal of Medicine*. 1998; 338(13):853-60.
16. Becker S. Liver toxicity in epidemiological cohorts. *Clinical Infectious Diseases*. 2004; 38(2):S49-S55.
17. Bonacini M. Liver injury during highly active antiretroviral therapy: The effect of hepatitis C coinfection. *Clinical Infectious Diseases*. 2004;38(Supplement_2):S104-S8.
18. Shiferaw MB, Tulu KT, Zegeye AM, Wubante AA. Liver enzymes abnormalities among highly active antiretroviral therapy experienced and HAART naïve HIV-1 infected patients at Debre Tabor Hospital, North West Ethiopia: A comparative cross-sectional study. *AIDS research and treatment*. 2016;2016.
19. Owiredu DWK, Quaye L, Amidu N. Oxidative stress and Dyslipidaemia among Ghanaian HAART-naïve HIV patients and those on HAART. *West African Journal of Pharmacy*. 2011;22(1).
20. Pol S, Lebra P, Vallet-Pichard A. HIV infection and hepatic enzyme abnormalities: Intricacies of the pathogenic mechanisms. *Clinical Infectious Diseases*. 2004;38(2):S65-S72.
21. Cote HC, Brumme ZL, Craib KJ, Alexander CS, Wynhoven B, Ting L, et al. Changes in mitochondrial DNA as a marker of nucleoside toxicity in HIV-infected patients. *New England Journal of Medicine*. 2002;346(11):811-20.
22. Sulkowski MS. Drug-induced liver injury associated with antiretroviral therapy that includes HIV-1 protease inhibitors. *Clinical Infectious Diseases*. 2004;38(2):S90-S7.
23. Lemoine M, Serfaty L, Capeau J. From nonalcoholic fatty liver to nonalcoholic steatohepatitis and cirrhosis in HIV-infected patients: Diagnosis and management. *Current Opinion in Infectious Diseases*. 2012;25(1):10-6.
24. Kontorinis N, Dieterich D. Hepatotoxicity of antiretroviral therapy. *AIDS Reviews*. 2003; 5(1):36-43.

25. Birkus G, Hitchcock MJ, Cihlar T. Assessment of mitochondrial toxicity in human cells treated with tenofovir: comparison with other nucleoside reverse transcriptase inhibitors. *Antimicrobial agents and chemotherapy*. 2002;46(3): 716-23.
26. Sulkowski MS, Thomas DL, Chaisson RE, Moore RD. Hepatotoxicity associated with antiretroviral therapy in adults infected with human immunodeficiency virus and the role of hepatitis C or B virus infection. *Jama*. 2000;283(1):74-80.
27. Flexner C. HIV-protease inhibitors. *New England Journal of Medicine*. 1998; 338(18):1281-93.
28. Iisu A, Granneman G, Bertz R. Ritonavir: Clinical pharmacokinetics and interactions with other anti-HIV agents. *Clin Pharmacokinet*. 1998;35:275-91.
29. Cameron W, Japour AJ, Xu Y, Hsu A, Mellors J, Farthing C et al. Ritonavir and saquinavir combination therapy for the treatment of HIV infection. *Aids*. 1999; 13(2):213-24.
30. Zucker SD, Qin X, Rouster SD, Yu F, Green RM, Keshavan P et al. Mechanism of indinavir-induced hyperbilirubinemia. *Proceedings of the National Academy of Sciences*. 2001;98(22):12671-6.
31. Rossen RD, Butler WT, Waldman RH, Alford RH, Hornick RB, Togo Y, et al. The proteins in nasal secretion: II. A longitudinal study of IgA and neutralizing antibody levels in nasal washings from men infected with influenza virus. *Jama*. 1970;211(7):1157-61.
32. Stiehm ER, Fudenberg HH. Serum levels of immune globulins in health and disease: a survey. *Pediatrics*. 1966;37(5):715-27.
33. Ngala RA, Opoku D, Asare G. Effects of HIV infection and highly active antiretroviral therapy (HAART) on the liver of HIV patients. *Trends in Medical Research*. 2015;10(1):1-11.
34. Rodriguez-Barradas MC, Alexandraki I, Nazir T, Foltzer M, Musher DM, Brown S, et al. Response of human immunodeficiency virus–infected patients receiving highly active antiretroviral therapy to vaccination with 23-valent pneumococcal polysaccharide vaccine. *Clinical Infectious Diseases*. 2003;37(3):438-47.
35. Picker LJ. Immunopathogenesis of acute AIDS virus infection. *Current Opinion in Immunology*. 2006;18(4):399-405.
36. Mahato SK, Kumar V, Jagmohankumar, Kumar A. Assessment of liver function Test in AIDS patients taking HAART At Rims Art Centre, Ranchi, Jharkhand, India. *IOSR Journal of Dental and Medical Sciences (IOSR-JDMS)*. 2016;15(12):59-61.
37. Ayelagbe O, Akerele O, Onuegbu A, Oparinde D. Drug hepatotoxicity in HIV patients on highly active antiretroviral therapy [HAART] in Southwest Nigeria. *IOSR-JDMS*. 2014;13(5):67-70.
38. Soriano V, Puoti M, Garcia-Gasco P, Rockstroh JK, Benhamou Y, Barreiro P, et al. Antiretroviral drugs and liver injury. *Aids*. 2008;22(1):1-13.
39. Betjes MG, Verhagen DW. Stable improvement of renal function after initiation of highly active anti-retroviral therapy in patients with HIV-1-associated nephropathy. *Nephrology Dialysis Transplantation*. 2002;17(10):1836-9.
40. Schwartz EJ, Cara A, Snoeck H, Ross MD, Sunamoto M, Reiser J, et al. Human immunodeficiency virus-1 induces loss of contact inhibition in podocytes. *Journal of the American Society of Nephrology*. 2001; 12(8):1677-84.
41. Shahinian V, Rajaraman S, Borucki M, Grady J, Hollander WM, Ahuja TS. Prevalence of HIV-associated nephropathy in autopsies of HIV-infected patients. *American Journal of Kidney Diseases*. 2000;35(5):884-8.
42. Kimmel PL. The nephropathies of HIV infection: Pathogenesis and treatment. *Current opinion in nephrology and hypertension*. 2000;9(2):117-22.

© 2020 Bani et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:
<http://www.sdiarticle4.com/review-history/59518>